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Novel synthetic receptors based on *para*-amino-pyridine ligands coupled to *p-tert*-butylcalix[4]arene via amino-acid spacers

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Abstract—The synthesis of a series novel receptors based on *p-tert*-butylcalix[4]arene tetra-substituted at the lower rim by pyridinyl derivatives of the amino acids, glycine, alanine and lysine is described. © 2001 Elsevier Science Ltd. All rights reserved.

The relative rigidity of various supramolecular systems makes them interesting candidates for the introduction of spatially differentiated coordinating systems, i.e. as di- or polytopic molecular receptors.¹ There has recently been increasing interest in the use of the calix[4]arene skeleton for the development of this type of multiple coordination-centre molecule.² The synthetic strategy behind such development involves coupling, generally at the lower face of a calix[4]arene, a group capable of coordinating either a metal, for example the porphyrin units used by Shinkaï,³ or for anions, the use of pyridinium units by Beer.⁴ Examples of ditopic complexation including the selective complexation of sodium phosphates,⁵ chloride or bromide⁶ and

caesium chloride⁷ were developed by Reinhoudt. The choice of the spacer unit between the calix[4]arene and the complexation site is a key factor in controlling both the size of the complexation centre and the selectivity in ditopic complexation. Similarly substitution along the spacer chain will influence the preorganisation at the complexation centre.

In this paper we report the synthesis of a series of pyridinium groups attached to calix[4]arene via aminoacid derived spacer arms. By using glycine (Gly) and alanine (Ala) we have been capable of varying the stereochemistry along the spacer and introducing chirality at this point. The use of N- ε -lysine (Lys) based



Scheme 1. Synthesis of pyridinium derivatives: (i) DCC, HOBT, DIEA, DMF; (ii) HClg; (iii) H2, Pd/C, MeOH, CH3COOH.

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systems allows variation in the length of the spacer as compared to the other two systems. The pyridine derivatives 4a, 4b and 4c were synthesised from the 4-aminopyridine 1 in two steps (Scheme 1). 4-Aminopyridine 1 (18 mmol) was condensed with the N-protected amino-acid derivatives *t*-butyloxycarbonyl-glycine (Boc-Gly-OH) 2a, t-butyloxycarbonyl-alanine (Boc-Ala-OH) **2b** and *t*-butyloxycarbonyl-lysine-*N*- ε -benzyloxycarbonyl (Boc-Lys(Z)-OH) 2c (15 mmol) in dry DMF (250 mL) in the presence of 1,3-dicyclohexylcarbodiimide (DCC, 18 mmol), hydroxy-benzotriazole (HOBT, 18 mmol) and diisopropylethylamine (DIEA, 21 mmol) for 3 h at 0°C followed by 2 h at 20°C. After elimination of dicyclohexylurea (DCU), the solvent was eliminated and the crude product purified by crystallisation from acetone for $3a^8$ (44% yield) and toluene for $3b^8$ (43% yield). Purification by chromatography on silica gel (CHCl₃: acetone 6:4) gave 3c in 77% yield.⁸ The derivatives 4a (95% yield) and 4b (97% yield)⁹ were obtained after complete deprotection by gaseous HCl in The benzyloxycarbonyl group (Z) was CH₂Cl₂. 3c by hydrogenolysis removed from in MeOH:CH₃COOH (1 M aqueous) 20:3 in the presence of the Pd/C 10% to yield 4c (91% yield).⁹

Calix[4]arene derivatives **6a**, **6b** (87 and 40% yield, respectively) and 7¹⁰ were synthesised by condensation at 25°C of the tetra-succinimidoyl activated ester **5**¹¹ with the pyridinium derivatives **4a**, **4b** or **4c** in CH₂Cl₂ in the presence of DIEA (Scheme 2). The desired compound **7**, for the lysine system, was obtained by deprotection of the *N*- α -Boc protecting group by means of gaseous HCl/CH₂Cl₂ treatment. The dichloride salt **7** is obtained in 73% yield. Electrospray mass spectrometry (positive mode) clearly demonstrates that **6a**, **6b** and **7** are tetrasubstituted derivatives (*m*/*z* 1414 [**6a**+H]⁺, 1492 [**6b**+Na]⁺ and 1699 [**7**+H]⁺ and that under-substituted derivatives are absent. The presence of mono, di and tri-charged species for each compound is to be noted.

¹H NMR shows the highly symmetrical nature of **6a** and **7**. The cone conformation¹² is confirmed by the presence of an AB (J=13.4 Hz) system characteristic of methylene bridges (ArCH₂Ar) and by the position of the carbon atoms of the methylene bridges in the ¹³C NMR: 31.52, 29.97 and 31.76 ppm for **6a**, **6b** and **7**, respectively. The presence of a stereogenic carbon near



Scheme 2. Synthesis of calixarenic ligand. (i) 4a or 4b, DIEA, CH_2Cl_2 , 25°C, 5 days; (ii) 1. 4c, DIEA, CH_2Cl_2 , 25°C, 5 days; 2. HCl_g , CH_2Cl_2 , 25°C, 2 hours.

the calixarene rim leads to structural modifications for **6b**, which are shown by an expanded AB system of the methylene diastereotopic protons (3.95-4.65 ppm) and by shielding at 6.54 ppm of the amide proton doublet (8.54 and 8.26 ppm for 6a and 7). There would therefore seem to be no hydrogen bonds for **6b** at the level of the crown constituted by the four NHCO, as has been observed in other works.¹³ This non-existence of hydrogen bonding leads to the unshielding of the CHa (5.21 ppm) compared to the CH α of **6a** (4.03 ppm) and of 7 (4.33 ppm). The aromatic calixarene protons of 6b are also disturbed and are differentiated in the form of two singlets at 7.11 and 7.14 ppm each of intensity 4, whereas one singlet alone of intensity 8 is observed for 6a (6.88 ppm) and for 7 (6.79 ppm). All these elements indicate a distortion of the calixarene cavity induced by the pro-helical structure due to the stereogenic carbons of 6b. A similar result has been observed in previous work.¹¹ However, for 7 the stereogenic carbon is too distant from the calix[4]arene cavity to give rise to this effect.

Transition metal complexation studies where the geometry of the coordination sphere at the metal centre is of square–planar type are in progress. The grafting four 4-amino-pyridine ligands on a calix[4]arene skeleton via amino-acid spacers leads to novel ditopic receptors and presents a potential route to porous organic structures.

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for $[M+H]^+$. **3b**: mp 169°C, ¹H NMR (DMSO-*d*₆ 300 MHz): δ (ppm) 1.25 (d, 3H, CH₃), 1.37 (s, 9H, Boc), 4.09 (t, 1H, H α), 7.20 (d, 1H, NHBoc), 7.57 (d, 2H, Hpy), 8.42 (d, 2H, Hpy), 10.33 (s, 1H, NHCO); ES–MS (*m*/*z*): 266 for $[M+H]^+$. **3c**: mp 63°C; ¹H NMR (CDCl₃ 300 MHz): δ (ppm) 1.45 (s, 9H, Boc), 1.55–1.96 (m, 6H, CH₂), 3.21 (q, *J*=6.6 Hz, 2H, CH₂ β), 4.19 (m, 1H, H α), 4.95 (t, *J*=6.3 Hz, 1H, NH(Z)), 5.10 (s, 2H, CH₂ Φ), 5.34 (d, *J*=7.7 Hz, 1H, NHBoc), 7.35–7.36 (m, 5H, H-Ar), 7.44 (d, *J*=6.2 Hz, 2H, Hpy), 8.44 (d, *J*=6.2 Hz, 2H, Hpy), 9.05 (s, 1H, NH-C=O); ES–MS (*m*/*z*): 457 for [M+H]⁺.

- Selected spectra data for 4a: ¹H NMR (DMSO-d₆+D₂O 300 MHz): δ (ppm) 3.92 (s, 2H, CH₂), 8.03 (d, J=7.35, 2H, Hpy), 8.64 (d, J=7.35, 2H, Hpy). 4b: (D₂O 300 MHz): δ (ppm) 1.66 (d, J=7.35 Hz, 3H, CH₃), 4.35 (q, J=7.35 Hz, 1H, CHα), 8.15 (d, J=7.35 Hz, 2H, Hpy), 8.63 (d, J=7.35 Hz, 2H, Hpy). 4c: (DMSO-d₆, 300 MHz): δ (ppm) 1.27 (m, 2H, CH₂), 1.37 (s, 9H, Boc), 1.54–1.65 (m, 4H, CH₂), 1.87 (s, 6H, CH₃COO⁻), 2.75 (m, 2H, CH₂), 4.06 (m, 1H, Hα), 7.18 (d, J=7.5 Hz, 1H, NHβ), 7.61 (d, J=5.9 Hz, 2H, Hpy), 8.41 (d, J=6.2 Hz, 2H, Hpy), 10.75 (s, 1H, NHpy); ES–MS(+) (m/z): 323 for [M-2CH₃COOH+H]⁺.
- 10. Selected spectra data for **6a**: mp 235°C, IR (KBr) 1677 (CO(NH)), 3334 (NH); UV (CH₂Cl₂): λ 242 nm (ϵ = 900000);¹H NMR (DMSO- d_6 300 MHz): δ (ppm) 1.06 (s, 36H, 'Bu), 3.25–4.57 (AB, J_{AB} =11.86 Hz, 8H, Ar-CH₂-Ar), 4.03 (d, J=4.41 Hz, 8H, CH₂ α), 4.61 (s, 8H, OCH₂), 6.88 (s, 8H, H-Ar), 7.48 (d, J=5.14 Hz, 8H, Hpy), 8.34 (d, J=3.68 Hz, 8H, Hpy), 8.55 (t broad, 4H, NH), 10.38 (s, 4H, NHpy); ¹³C NMR: δ (ppm) 31.52, (Ar-CH₂-Ar), 31.96 (Me, 'Bu), 34.46 (C, 'Bu), 43.62 (C α), 74.78 (OCH₂), 114.05 (H-C_{py}), 126.30 (3, 5-Ar), 133.65, 145.64, 146.22, 154.11 (1,2,4,6-Ar, 4-Py), 151.09 (H-C_{py}), 169.65, 170.67 (CONH); ES–MS (+) (m/z): 1414 for [M+H]⁺, 1436 for [M+Na]⁺. Anal. calcd for C₈₀H₉₂O₁₂N₁₂+Na⁺+ 4H₂O: C, 63.67; H, 6.68; N, 11.14; O, 16.97. Found C, 63.83; H, 6.70; N, 10.80.
 - 6b: mp 240°C, IR (KBr) 1675.8 (CO(NH)), 3331.6 (NH); UV (CH₂Cl₂): λ 242 nm (ϵ =900000);¹H NMR (DMSO d_6 300 MHz): δ (ppm) 1.15 (s, 36H, 'Bu), 1.18 (d, J=6.88 Hz, 12H, CH₃), 3.33–4.37 (AB, J_{AB}=11.86 Hz, 8H, Ar-CH₂-Ar), 3.95–4.65 (AB, J_{AB}=13.39 Hz, 8H, OCH₂), 5.21 (m, J = 6.51 Hz, 4H, CH_{α}), 6.54 (d, J = 5.74 Hz, 4H, NH-Ca), 7.11 (s, 4H, H-Ar), 7.14 (s, 4H, H-Ar), 7.88 (dd, $J_1 = 4.97$ Hz, $J_2 = 1.53$ Hz, 8H, H-Py), 8.49 (dd, $J_1 = 4.98$ Hz, $J_2 = 1.53$ Hz, 8H, H-Py), 10.34 (s, 1H, NH-Py); ¹³C NMR 20.60 (CH₃-Cα), 29.97 (Ar-CH₂-Ar), 31.67 (Me, ^tBu), 34.55 (C, ^tBu), 50.57 (Ca), 74.81 (OCH₂), 113.7 (H-C_{pv}), 125.90, 126.27 (3, 5-Ar), 134.58, 135.15, 145.83, 148.41, 150.27 ((1,2,4,6-Ar, 4-Py), 151.53 (H-C_{py}), 168.83, 172.96 (CONH); ES-MS (+) [M+Na]⁺ 1492. Anal. calcd for $C_{84}H_{100}O_{12}N_{12}+H_2O+Na^+$: C, 66.76; H, 6.81; N, 11.13; O, 13.77. Found: C, 66.69; H, 6.98; N, 11.19. 7: mp 239°C, IR (KBr) 1678 (CO(NH)), 3330 (NH); UV $(CH_2Cl_2) \lambda 242 \ (\varepsilon = 880000); {}^{1}H NMR \ (DMSO-d_6 500)$ MHz): δ (ppm) 1.03 (s, 36H, ^tBu), 1.44 (s brd, 16H, $CH_2\gamma + CH_2\delta$), 1.97 (s brd, 8H, $CH_2\beta$), 3.11 (s brd, 8H, CH₂ ϵ), 3.18–4.48 (AB, J_{AB} =13.6 Hz, 8H, ArCH₂Ar), 4.33 (s brd, 4H, CHa), 4.42 (s, 8H, OCH₂), 6.79 (s, 8H, HAr), 8.26 (d, J=6.6 Hz, 8H, Hpy), 8.41 (s brd, 4H, NHCH₂ ϵ), 8.73 (s brd, 12H, NH₃⁺), 8.79 (d, J=6.3, 8H, Hpy), 12.87 (s large, 4H, NH); ¹³C NMR 22.50, 29.47,

31.20 (CH₂ β , γ , δ), 31.76 (ArCH₂Ar), 31.96 (Me), 34.38 (C'Bu), 38.98 (CH₂ ϵ), 54.18 (CH α), 74.76 (OCH₂), 115.67, 143.68 (2,3,5,6-py), 126.11 (3,5-Ar), 133.59, 145.20, 152.94, 153.82 (1,2,4,6-Ar+4-py), 169.75, 171.12 (CONH); ES–MS (+) [M+H]⁺ 1699. Anal. calcd for C₉₆H₁₃₆O₁₂N₁₆Cl₈+6H₂O+2Na⁺: C, 53.73; H, 6.96; N, 10.45. Found C, 53.85; H, 7.04; N, 10.4.

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